

0960-894X(94)00421-8

POTENT TRIAZOLINONE-BASED ANGIOTENSIN II RECEPTOR ANTAGONISTS WITH EQUIVALENT AFFINITY FOR BOTH THE AT₁ AND AT₂ SUBTYPES¹

L. L. Chang,* W. T. Ashton, K. L. Flanagan, R. A. Rivero, T.-B. Chen,† S. S. O'Malley,† G. J. Zingaro,† S. D. Kivlighn,† P. K. S. Siegl,† V. J. Lotti,† R. S. L. Chang,† and W. J. Greenlee

Merck Research Laboratories, Rahway, NJ 07065 and †West Point, PA 19486

Abstract: A series of subnanomolar (IC $_{50}$) triazolinone-based AT $_{1}$ /AT $_{2}$ -balanced AII antagonists has been identified. The 70–240-fold gain in AT $_{2}$ activity relative to prototype compounds was achieved by the introduction of a 5-acylamino group on the N $_{2}$ -aryl moiety and the addition of (3-F-5'-Pr)biphenyl substituents on 4. These analogues exhibited AT $_{2}$ /AT $_{1}$ IC $_{50}$ ratios of \leq 1 in multiple assay systems including human adrenal.

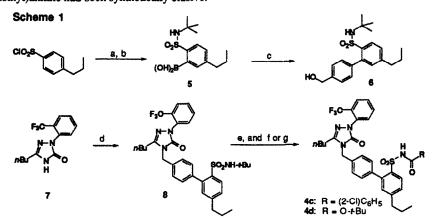
Angiotensin II (AII) is the principal effector hormone of the renin-angiotensin system (RAS).² This octapeptide has equivalent affinity for both of the major subtypes of the AII receptor (AT₁ and AT₂).³ The AT₁ receptor is crucial in the regulation of blood pressure and electrolyte homeostasis, and is associated with most of the known physiological effects of AII.³ The investigative drug Cozaar[®] (losartan, DuP 753, MK-954, 1) is a nonpeptide AT₁-selective antihypertensive agent.⁴ The physiological role of the AT₂ receptor has yet to be clearly defined although it has been associated with the regulation of renal function,⁵ and may be involved in restenosis following vascular injury,⁶ in wound healing,⁷ and in cardiac fibroblast collagen synthesis.⁸ In addition, it has been implicated in various cell differentiation and cell proliferation processes.^{3,9} Recently, high affinity nonpeptide AT₂-selective ligands have been described also.¹⁰ The administration of an AT₁-selective AII antagonist results in an increase in the plasma levels of AII.¹¹ The effect of prolonged stimulation of AT₂ receptors by elevated levels of AII is not known. Dual-action AII antagonists capable of simultaneous blockade of both receptor subtypes (AT₁/AT₂-balanced AII antagonists) would be useful as pharmacological tools and could have advantages as therapeutic agents over AT₁-selective compounds.

Many dual-action peptide ligands for the AII receptor are known but have unsatisfactory pharmacokinetic properties, and are of limited pharmacological value.^{3,12} The discovery of a series of high affinity AT_1/AT_2 -balanced (N-alkyl-N-acyl)aminoquinazolinone tetrazoles such as 2 (L-159,689),¹³ paved the way towards the identification of other dual-action AII antagonists.¹⁴ In the triazolinone series, starting from a benzoylsulfonamide (4, $R^1 = CF_3$, $R^2 = R^3 = R^4 = H$, $R^5 = C_6H_5$, L-159,913) with an AT_2 IC50 value of 300 nM (rat midbrain),¹⁵ judicious replacement of the N-substituent of the sulfonamide, and the addition of a 5-

acylamino group on the N^2 -aryl moiety resulted in compound 3 (L-163,007) which had an AT₂ IC₅₀ value of 1 nM and an AT₂/AT₁ IC₅₀ ratio of 3 (rat midbrain/rabbit aorta). Other approaches to enhance AT₂ binding affinity include: (1) replacement of the terminal phenyl ring of the biphenyl moiety by a 5-alkyl-substituted thienyl group as shown in a series of imidazopyridine-based compounds, ¹⁶ and (2) the addition of a 3-fluoro substituent on the biphenyl moiety in a series of imidazole-based acyl (or related) sulfonamides, demonstrated by the DuPont Merck group. ¹⁷ Modifying triazolinone biphenylsulfonamides 4a and 4b which had modest AT₂ binding affinity ¹⁸ (see Table 1), we have prepared and evaluated compounds 4c-m in attempts to obtain structurally diverse, potent, and fully balanced AII antagonists which show oral activity in animal models. To ensure equivalent coverage of both receptors under physiological conditions, we looked for compounds with AT₂/AT₁ IC₅₀ ratios of ≤ 1 in three pairs of tissue preparations (rat midbrain/rabbit aorta, rat adrenal, and human adrenal).

Chemistry

Two routes were used to prepare these compounds, depending on whether R^3 in 4 is H or F. For compounds with $R^3 = H$ (4c,d,h,i)¹⁹, the requisite biarylmethyl side chain was appended onto the 4-unsubstituted triazolinone under Mitsunobu conditions. The synthesis for compounds 4c,d is shown in Scheme 1. 4-n-Propylbenzenesulfonyl chloride was made into its N-t-butylsulfonamide, which was deprotonated by excess n-butyllithium and then treated with triisopropyl borate to furnish the boronic acid 5.20 Palladium(0)-catalyzed biaryl coupling between 4-bromobenzyl alcohol and 5 provided the corresponding biphenylmethanol 6.21a Alkylation of the triazolinone 718 using 6, triphenylphosphine, and diisopropyl azodicarboxylate²² provided the intermediate 8. Removal of the t-butyl group followed by acylation or alkoxycarbonylation according to previously described procedures^{1,18} gave target compounds 4c,d. Analogues 4h-m have a chloro instead of a trifluoromethyl group at R^1 since 2-chloro-5-nitrophenylhydrazine, required for the synthesis of the key intermediate 9 for these derivatives, was readily available from the corresponding aniline while 5-nitro-2-(trifluoromethyl)aniline had been synthetically elusive.¹



a: (i) \pm BuNH₂/CH₂Cl₂ 0°C- rt, (ii) NaOH. b: (i) 2.5 eq. π BuLl/THF, -40°C- rt, (ii) (\pm PrO)₃B, 0°C- rt (iii) aq. HCl, 90% for a,b. c: 3 eq. (4-Br)C₆H₃C H₂OH, 3 mol% Pd(PPh₃)₄, NaOH, toluene/EtOH, 100°C, 3h, 62%. d: 6, P(Ph)₃, DIAD/THF, -15°C- rt, 66%. e: TFA, rt, 73%. f: (2-Cl)C₆H₅C O₂H, Im₂CO, DBU/THF, 86%. g; NaH, (BOC)₂O/THF, 95%.

The synthesis for compounds 4k,l, shown in Scheme 2, is typical of the route taken to prepare analogues with a 3-fluoro substituent on the biphenyl moiety (4e-g, j-m). Alkylation of the anion of the N⁴-unsubstituted

triazolinone 9^1 by 4-bromo-2-fluorobenzyl bromide provided the intermediate 10. This compound underwent Pd(0)-catalyzed cross coupling reaction with boronic acid 5 to provide $11,^{21}$ containing the 3-fluoro-5'-propylbiaryl sulfonamide moiety. The nitro group on the N^2 -aryl was reduced by stannous chloride and then acylated using acetyl chloride. Subsequent removal of the t-butyl group provided the free sulfonamide $12,^{1,18}$ Acylation or alkoxycarbonylation furnished the desired analogues $4k,l.^{1,18}$

Scheme 2 CI NO2 N-NO2 N-NO2

a: NaH/DMF, (4-Br,2-F)C $_{9}$ H $_{3}$ CH $_{2}$ Br, 50'-90'C, 79%. b: 2 eq. 5, 5 mol% Pd(PPh $_{3}$) $_{4}$, 4 eq. NaOH, toluene/EtOH, 90'C, 6 h, 68%. c: H $_{2}$ /PtO $_{2}$, EtOH, 78%. d: DMAP, pyridine, ClCOMe, 88%. e: TFA, 88%. f: (2-F)C $_{9}$ H $_{4}$ CO $_{2}$ H, Im $_{2}$ CO, DBU/THF, 56%. g: NaH, (BOC) $_{2}$ O/THF, 72%.

Biological Results and Discussion

The ability of compounds 4c-m to block competitively the specific binding of the radioligand $^{125}I[Sar^1,Ile^8]AII$ to a rabbit aorta (for AT_1 receptor) and a rat midbrain (for AT_2 receptor) membrane preparation was assessed as previously described. 23 Compounds which showed AT_2/AT_1 IC_{50} ratios of ≤ 5 were further evaluated in the rat and/or human adrenal AT_1 and AT_2 receptor tissue preparations. 24 Multiple runs of the assays were conducted for each key compound to ensure consistency in the IC_{50} values obtained. For simplicity, Table 1 shows only data from the aorta/midbrain and human adrenal assays. In general, these ligands were equally potent or, slightly more active at the rat adrenal AT_1 and AT_2 receptors compared to the aorta/midbrain combination. The AT_2/AT_1 IC_{50} ratios were generally very similar for these pairs.

Data from analogues 4c,d show that, compared with 4a,b, the added 5'-propyl group provided a 2-3-fold increase in the AT₂ binding affinity but resulted in a 10-20-fold loss in AT₁ potency. Compound 4c, with a 2-chlorobenzoylsulfonamide moiety, was less adversely affected on AT₁ than the sulfonylcarbamate 4d. This effect of the 5'-alkyl group in increasing the AT₂ binding affinity at the expense of AT₁ potency was also observed in other heterocyclic series. Analogue 4d was a balanced compound from the aorta/midbrain assays but was not examined further because of insufficient aorta AT₁ potency and poor IC₅₀ values from the human adrenal assays. Compounds 4e,f contain a 3-fluoro substituent on the biarylmethyl moiety. Comparing these with the unsubstituted acylsulfonamide 4a and sulfonylcarbamate 4b, a 3-fold increase in AT₂ binding affinity was achieved while maintaining subnanomolar AT₁ potency. From these data, we inferred that 4g, an

acylsulfonamide containing both the 3-F and 5'-Pr substituents, could be a balanced compound. Indeed, 4g was balanced in all pairs of tissue preparations (rat adrenal AT_{1IC50}=1.5 nM, AT_{2IC50}=1.1 nM). High human adrenal IC₅₀ values, however, precluded further interest.

TABLE 1. IN VITRO SAR OF VARIOUS N^2 -ARYL TRIAZOLINONE BIPHENYLSULFONAMIDES FOR THE AT₁ AND AT₂ RECEPTOR SUBTYPES OF AII

Cpd	_	R2	_R3_	R ⁴ _	R5	Rabbit Aorta/Rat Midbraina			Human Adrenal		
						IC ₅₀ .	(nM) AT ₂	AT ₂ /AT ₁ IC ₅₀ Ratio	IC50 (I		AT ₂ /AT ₁ IC ₅₀ Ratio
4ab	CF ₃	н	Н	н	(2-CI)Ph	0.11	36	300			
4b ^b	CF ₃	н	н	н	O-t-Bu	0.45	17	39			
4 c	CF ₃	н	н	n-Pr	(2-CI)Ph	1.3	14	11			
4 d	CF ₃	н	н	n-Pr	O-t-Bu	10	8.1	0.8	150	270	1.8
4 e	CF ₃	н	F	Н	(2-CI)Ph	0.15	12	80			
41	CF ₃	н	F	н	O-t-Bu	0.31	4.2	14			
4 g	CF ₃	н	F	n-Pr	(2-CI)Ph	3.9	3.5	0.9	110	140	1.3
4 h	CI	NHCO-n-Bu	н	Et	(2-CI)Ph	1.0	4.9	4.9			
41	CI	NHCOEt	н	Et	(2-CI)Ph	0.25	3.5	14			
41	CI	NHCOMe	F	n-Pr	(2-CI)Ph	0.90	0.15	0.2	16 (0.489)	17	1.1
4 k	CI	NHCOM.	F	n-Pr	(2-F)Ph	0.62	0.15	0.2	5.9	2.4	0.4
41	CI	NHCOMe	F	n-Pr	O- <i>t</i> -Bu	0.84	0.24	0.3	14 (0.82%)	12	0.9
4 m	CI	NHCOMe	F	n-Pr	O-Et	1.4	0.10	0.07	15	2.6	0.2
4nd	CI	NHCO-n-Bu	н	н	(2-CI)Ph	0.16	1.6	10			
40 ^d	CI	NHCOEt	н	н	(2-CI)Ph	0.17	2.5	15			
4pd	CI	NHCOMe	н	н	(2-CI)Ph	0.052	12	230			

 a For the rabbit aorta and rat midbrain binding assays, no bovine serum albumin (BSA) was added to the assay mixtures. For the human adrenal assays, 0.2% BSA was present in the assay mixtures unless otherwise indicated. b This compound was reported in reference 18. c Data from cloned human AT1 receptor (no BSA in assay mixture). d This compound was reported in reference 1.

Subsequently, we investigated compounds 4h, i which have a 5'-ethyl substituent and an amide moiety on the 5-position of the N²-aryl ring. The amide moiety is known to enhance AT₂ activity in the triazolinone series.¹ Compared to the 5'-propyl group, the 5'-ethyl group was expected to induce a smaller increase in AT₂ binding affinity but suffer less of a loss in AT₁ potency. Together, these modifications might result in a balanced compound more potent than 4d or 4g. Table 1 shows that, unfortunately, the 5'-ethyl group resulted in a decrease in AT₂ binding affinity in 4h, i compared to 4n, o, unsubstituted at the 5'-position, and the AT₂/AT₁ IC₅₀ ratio remained unsatisfactory in the 5'-ethyl analogues.

Next, we considered using a combination of an acetylamino group at the 5-position of the N^2 -aryl and 3-fluoro-5'-propylbiaryl substituents to enhance AT_2 potency. The expected loss in AT_1 potency owing to the 5'-propyl group was designed to be partially offset by the presence of the acetylamino group, which had previously provided an exceptionally potent AT_1 derivative (e.g., 4p vs. 4o).\(^1\) Four analogues were synthesized in this series, 4j-m. The first compound prepared, 4j (rat adrenal AT_{1IC50} =0.38 nM, AT_{2IC50} =0.11 nM), had

subnanomolar intrinsic potency at both receptors, superior in this respect compared to a close analogue 4g. In addition, 4j was balanced in all 3 sets of tissue preparations. The 2-fluorobenzoylsulfonamide 4k was slightly more favored at AT_1 , ¹⁸ providing a compound with AT_2/AT_1 IC₅₀ ratios consistently <1. The *t*-butyl sulfonyl-carbamate 4l (rat adrenal $AT_{1KC50}=0.70$ nM, $AT_{2KC50}=0.17$ nM) also met our criterion for a balanced compound (human adrenal AT_2/AT_1 IC₅₀ ratio ~1). The cloned human AT_1 receptor IC₅₀ values for 4j,l were 0.48 nM and 0.82 nM, respectively.²⁵ These data suggest an approximate equivalence in intrinsic potency between the rabbit aorta AT_1 and the human AT_1 receptors. Finally, the ethyl sulfonylcarbamate 4m showed a ~6–14-fold preference for the AT_2 receptor. Relative to the *t*-butyl group, the less bulky ethyl group was able to achieve greater AT_2 binding affinity but incurred a modest loss of AT_1 potency.

The inhibition of pressor response to exogenous AII challenges by 4j, was studied in conscious normotensive rats according to protocols described previously.²⁶ At 3 mg/kg i.v., 4j showed 71% peak inhibition with a duration of 5 h and 4l showed 41% peak inhibition. Neither 4j nor 4l was active orally at this dose. This contrasts dramatically with conscious rat data for compounds 4a, b and 3. For these compounds, which contain an acylsulfonamide or sulfonylcarbamate, with or without an acylamino moiety, excellent efficacy (generally >85% peak inhibition) and long duration of action (6 < t < 24 h) were observed at 1 mg/kg both i.v. and orally.^{1,18} The DuPont Merck AII group has shown that the 3-F substituent on the biaryl moiety is consistent with good oral activity in rats.¹⁷ Therefore, the present data imply that, in the triazolinone series, the 5'-propyl group on the biaryl (perhaps in conjunction with the 2-chloro substituent on the N²-aryl) appear to reduce in vivo efficacy and adversely affect oral activity in conscious rats.

In summary, we have described triazolinone-based dual AII antagonists which showed subnanomolar binding affinity at both receptor subtypes and met goals for AT₁/AT₂ balance in multiple tissue preparations, including human adrenal. One analogue showed modest AT₂ selectivity. The 70–240-fold gain in AT₂ activity seen in 4j-m compared to 4a,b was primarily achieved by a 5-acetylamino group at the N²-aryl ring of the triazolinone and 3-fluoro, 5'-propyl substituents on the biaryl moiety. This study assisted our efforts to achieve fully balanced and orally active triazolinone-based AII ligands, which will be disclosed in the near future.

Acknowledgments

We thank Dr. L. F. Colwell, Jr. and Ms. A. Bernick for mass spectral determinations.

References and Notes

- 1. Preceding paper in this series: Chang, L. L.; Ashton, W. T.; Flanagan, K. L.; Chen, T.-B.; O'Malley, S. S.; Zingaro, G. J.; Siegl, P. K. S; Kivlighn, S. D.; Lotti, V. J.; Chang, R. S. L.; Greenlee, W. J. J. Med. Chem. in press.
- 2. Vallotton, M. B. Trends Pharmacol. Sci. 1987, 8, 69.
- (a) Timmermans, P. B. M. W. M.; Wong, P. C.; Chiu, A. T.; Herblin, W. F.; Benfield, P.; Caraini, D. J.; Lee, R. J.; Wexler, R. R.; Saye, J. A. M.; Smith, R. D. Pharmacol. Rev. 1993, 45, 205. (b) Bottari, S. P.; de Gasparo, M.; Steckelings, U. M.; Levens, N. R. Front. Neuroendocrinol. 1993, 14, 123.
- (a) Duncia, J. V.; Carini, D. J.; Chiu, A. T.; Johnson, A. L.; Price, W. A.; Wong, P. C.; Wexler, R. R.; Timmermans, P. B. M. W. M. Med. Res. Rev. 1992, 12, 149. (b) Carini, D. J.; Duncia, J. V.; Aldrich, P. E.; Chiu, A. T.; Johnson, A. L.; Pierce, M. E.; Price, W. A.; Santella, J. B. III; Wells, G. J.; Wexler, R. R.; Wong, P. C.; Yoo, S.-E.; Timmermans, P. B. M. W. M. J. Med. Chem. 1991, 34, 2525. For reviews of AT₁-selective AII antagonists, see: (c) Murray, W. F. Chemtracts-Org. Chem. 1993, 6, 263. (d) Bühlmeyer, P. Curr. Opin. Ther. Pat. 1992, 1693. (e) Hodges, J. C.; Hamby, J. M.; Blankley, C. J. Drugs Future 1992, 17, 575.
- (a) Keiser, J. A.; Bjork, F. A.; Hodges, J. C.; Taylor, D. G., Jr. J. Pharmacol. Exp. Ther. 1992, 262, 1154.
 (b) Cogan, M. G.; Liu, F. Y.; Wong, P. C.; Timmermans, P. B. M. W. M. J. Pharmacol. Exp. Ther. 1991, 259, 687.
- (a) Janiak, P.; Pillon, A.; Prost, J.-F.; Vilaine, J.-P. Hypertension 1992, 20, 737.
 (b) Pratt, R. E.; Wang, D.; Hein, L.; Dzau, V. J. Hypertension, 1992, 20, 432.

- 7. Viswanathan, M.; Saavedra, J. Peptides, 1992, 13, 783.
- 8. Brilla, C. G.; Zhou, G.; Matsubara, L., Weber, K. T. J. Mol. Cell. Cardiol. 1994, 26, 809.
- (a) de Gasparo, M.; Whitebread, S.; Levens, N.; Ramjoué, H.-P.; Criscione, L.; Rogg, J.; Baum, H.-P.; Brechler, V.; Buehlmayer, P.; Wood, J. M.; Bottari, S. P. In Cellular and Molecular Biology of the Adrenal Cortex (Colloque INSERM, Vol. 222); Saez, J. M.; Brownie, A. C.; Capponi, A.; Chambaz, E. M.; Mantero, F., Eds.; J. Libbey Eurotext: Paris, 1992, pp 3-17. (b) Wong, P. C.; Hajj-ali, A. F. in Medicinal Chemistry of the Renin Angiotensin System, Timmermans, P. B. M. W. M.; Wexler, R. R.; Eds., Elsevier, New York, in press (1994).
- (a) Wu, M. T.; Ikeler, T. J.; Ashton, W. T.; Chang, R. S. L.; Lotti, V. J.; Greenlee, W. J. BioMed. Chem. Lett. 1993, 3, 2023.
 (b) Klutchko, S.; Hamby, J. M.; Hodges, J. C. BioMed. Chem. Lett., 1994, 4, 57.
- a) Christen, Y.; Waeber, B.; Nussberger, J.; Porchet, M.; Borland, R. M.; Lee, R. J.; Maggon, K.; Shum, L.; Timmermans, P. B. M. W. M.; Brunner, H. R. Circulation 1991, 83, 1333. (b) Goldberg, M. R.; Tanaka, W.; Barchowsky, A.; Bradstreet, T. E.; McCrea, J.; Lo, M.-W.; McWilliams, E. J.; Bjornsson, T. D. Hypertension 1993, 21, 704. (c) Abdelrahman, A. M.; Burrell, L. M.; Johnston, C. I. J. Hypertens. 1993, 11 (Suppl. 3) S23.
- 12. Streeten, D. H. P.; Anderson, G. H., Jr. In Clinical Pharmacology of Antihypertensive Drugs (Handbook of Hypertension, Vol. 5); Doyle, A. I., Ed.; Elsevier: Amsterdam, 1984, pp 264-271.
- de Laszlo, S. E.; Quagliato, C. S.; Greenlee, W. J.; Patchett, A. A.; Chang, R. S. L.; Lotti, V. J.; Chen, T.-B.; Scheck, S. A.; Faust, K. A.; Kivlighn, S. S.; Schorn, T. S.; Zingaro, G. J.; Siegl, P. K. S. J. Med. Chem. 1993, 36, 3207.
- (a) Glinka, T. W.; de Laszlo, S. E.; Siegl, P. K. S.; Chang, R. S.; Kivlighn, S. D.; Schorn, T. S. Faust, K. A.; Chen, T.-B.; Zingaro, G. J.; Lotti, V. J.; Greenlee, W. J. BioMed. Chem. Lett. 1994, 4, 81. (b) Mantlo, N. B.; Kim, D.; Ondeyka, D.; Chang, R. S. L.; Kivlighn, S. D.; Siegl, P. K. S.; Greenlee, W. J. BioMed. Chem. Lett. 1994, 4, 17.
- Chang, L. L.; Ashton, W. T.; Flanagan, K. L.; Naylor, E. M.; Chakravarty, P. K.; Patchett, A. A.; Greenlee, W. J.; Bendesky, R. J.; Chen, T.-B.; Faust, K. A.; Kling, P. J.; Schaffer, L. W.; Schorn, T. W.; Zingaro, G. J.; Chang, R. S. L.; Lotti, V. J.; Kivlighn, S. D.; Siegl, P. K. S. BioMed. Chem. Lett. 1994, 4, 115.
- 16. Kevin, N. J.; Rivero, R. A.; Greenlee, W. J.; Chang, R. S. L.; Chen, T. B. BioMed. Chem. Lett. 1994, 4, 189. A series of triazolinone-based substituted phenylthiophene benzoylsulfonamides has also been studied and showed SAR trends similar to those seen for the imidazopyridine-based compounds discussed in this reference: Rivero, R. A.; Greenlee, W. J.; Chang, R. S. L.; Chen, T.-B. Unpublished results.
- 17. We thank the DuPont Merck AII group for sharing pre-publication results on this finding: Quan, M. L.; Olson, R. E.; Carini, D. J.; Ellis, C. D.; Hillyer, G. L.; Lalka, G. K.; Liu, J.; VanAtten, M. K.; Chiu, A. T.; Wong, P. C.; Wexler, R. R.; Timmermans, P. B. M. W. M. BioMed. Chem. Lett. 1994, 4, 2011.
- Ashton, W. T.; Chang, L. L.; Flanagan, K. L.; Hutchins, S. M.; Naylor, E. M.; Chakravarty, P. K.; Patchett, A. A.; Greenlee, W. J.; Chen, T.-B.; Faust, K. A.; Chang, R. S. L.; Lotti, V. J.; Zingaro, G. J.; Schorn, T. W.; Siegl, P. K. S.; Kivlighn, S. D. J. Med. Chem. 1994, 37, 2808.
- 19. These and all other new compounds discussed were characterized by mp, mass spectrum (FAB), 300 MHz or 400 MHz ¹H-NMR, and elemental analyses (C, H, N; within ±0.4% of theoretical values) or high resolution mass spectrum.
- 20. Sharp, M. J.; Chen, W.; Snieckus, V. Tetrahedron Lett. 1987, 28, 5093.
- 21. (a) Miyaura, N.; Yanagi, T.; Suzuki, A. Synth. Commun. 1981, 11, 513. (b) This reaction is known to be capricious when the starting material contains a nitro group. In the synthesis of 11, decomposition (as observed by TLC) necessitated that the reaction be quenched before all of 10 was consumed (27% recovered). The yield quoted in Scheme 2 was based on the amount of 10 consumed.
- 22. Mitsunobu, O. Synthesis, 1981, 1.
- 23. (a) No bovine serum albumin (BSA) was added to the assay mixtures. The standard error (expressed as percent of mean) of the IC₅₀ measurement in this assay has been estimated to be <30%, based on the results of several standard compounds having 3 or more determinations. For detailed descriptions, see: (b) Chang, R. S. L.; Siegl, P. K. S.; Clineschmidt, B. V.; Mantlo, N. B.; Chakravarty, P. K.; Greenlee, W. J.; Patchett, A. A.; Lotti, V. J. J. Pharmacol. Exp. Ther. 1992, 262, 133. (c) Chang, R. S. L.; Lotti, V. J.; Chen, T. B.; Faust, K. A. Biochem. Biophys. Res. Commun. 1990, 171, 813.</p>
- 24. In order to achieve a satisfactory ratio of specific to nonspecific binding, it was necessary to add BSA (2 mg/mL) to the binding buffer for the human adrenal assays, but not for the others. Since both the AT₁ and AT₂ receptors are present in the rat and human adrenal tissues, IC₅₀ values on AT₁ and AT₂ were determined in the presence of 1 μM PD121981 (WL-19) or losartan to prevent binding of the radioligand to the AT₂ and AT₁ receptors, respectively. Otherwise, assay protocols were as described in 23b,c.
- 25. Human AT₁ receptors cloned in CHO cells were used in this assay. No BSA was present in the assay mixture.
- 26. At least two animals were evaluated in all cases. A ≥30% inhibition of the AII pressor response was considered significant in this assay. For details of experimental protocol, see: Siegl, P. K. S.; Chang, R. S. L.; Mantlo, N. B.; Chakravarty, P. K.; Ondeyka, D. L.; Greenlee, W. J.; Patchett, A. A.; Sweet, C. S.; Lotti, V. J. J. Pharmacol. Exp. Ther. 1992, 262, 139.